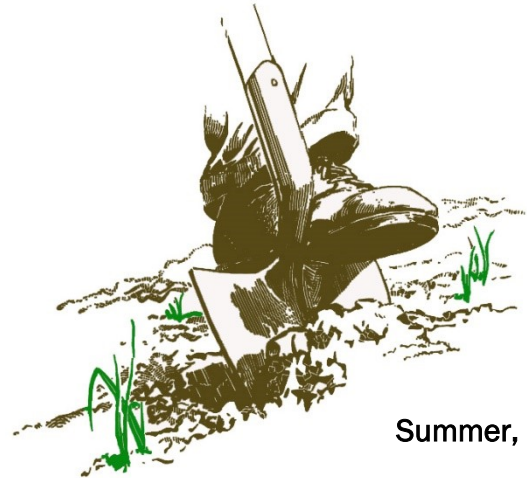


ON THE GROUND: TROUBLE SHOOTING SOIL AND CROP PRODUCTIVITY

Robert Walters¹, Dan Currie², Levi Walker², and Ruth Baldwin²



Overview

Optimizing productivity in row crops entails balancing the supply of air, water, and nutrients in the root zone using the principles of sound land husbandry. When the supply of essential nutrients cannot satisfy the demands of crop metabolism, the yield potential is discounted generally according to the [law of the minimum](#), wherein the most limiting essential element prevails until the final n^{th} element has been removed. Signs of crop deficiency may be exhibited via plant stunting, diurnal wilting, leaf discoloration, insect and disease damage, and low product quality and yield. Identifying yield-limiting factors requires measuring critical soil and plant index properties. Results from such tests may reveal inherent soil limitations, or indicate changes in existing crop management practices to sustain long term productivity.

Objectives

This 8-week project introduced agriculture interns at the Center for Environmental Farming Systems in Goldsboro, N.C. to the art and science agronomic problem solving. Together, interns were expected to: (1) develop an ability to observe, identify, and differentiate visual indicators of crop health and productivity; (2) acquire skill in measuring soil and plant properties and interpreting the results; and (3) draw conclusions and make recommendations based on soil-plant-environmental data.

This investigation focused on several non-contiguous areas of poor crop growth in the SARE long-term cropping systems crop-animal rotation plot # 20 (Figure 1). Symptoms of stunting, yellowing, acute toxicity, or death had been evident in corn and cotton over the past two seasons with no explanation (Figure 2).

The team's objectives were:

- Identify the cause of crop stunting and acute toxicity in SARE crop animal rotation plot #20.
- Make specific recommendations to correct these problems.
- Propose soil and crop management strategies to enhance long-term productivity.



Figure 1. SARE crop/animal (C/A) plot 20. Red arrow points to approximate study location.

¹ Research Specialist and author, Department of Soil Science

² Agriculture intern, Center for Environmental Farming Systems

Methods

Site Description

Information about the site physical setting, as well as present and past agronomic practices, is essential for providing a backdrop to initiate an investigation. Therefore, interns were asked to retrieve information about the study site using published USDA Soil Surveys and existing farm records. This is summarized in Table 1.

Sampling

We used the method of paired sampling to analyze the problem. Representative soil samples were collected in pairs from the problem site (Pr site) and from one healthy, non-problem site (non-Pr site) in SARE crop-animal rotation plot 20 (CAR-20). Within each sample site, two row positions were evaluated: (1) non-traffic, approximately 15 cm from the plant row; and (2) trafficked interrow. A total of twelve soil cores were collected from both row positions. A custom drive hammer tool fabricated to accept a standard 3.2 cm diameter steel Giddings drive tube was used to recover undisturbed soil cores down to 45 cm deep (Figure 3). Cores were sectioned in 0-15, 15-30, 30-45 cm deep increments and composited. For each depth increment, sub-samples were reserved for laboratory testing. Soil physical properties were measured at both sites and at each of two row positions to 15 cm deep. Subsurface soil samples were obtained by boring 150 cm deep using a 7.62 cm diameter bucket auger. The soil's surficial and subsurface characteristics were visually examined and described. In addition, twelve corn leaf samples composed of the most recently matured (MRM) leaf were harvested and submitted to the North Carolina Department of Agriculture (NCDA) labs for nutrient analysis.



Figure 2. Ground view of the problem site (Pr site) where this investigation was conducted. Photo shows a progression of crop symptoms from plant death (foreground), stunting and discoloration (middle), to healthy corn (background).

Table 1. Site description

Location	SARE long-term crop/animal rotation treatment plot 20		
Landowner	North Carolina Department of Agriculture & Consumer Services Center for Environmental Farming Systems, Cherry Farm Research Unit Goldsboro, Wayne County, NC		
Physiographic Province	Coastal Plain		
Physiographic Area	Neuse river basin		
Topographic Orientation	Soil Information		
Elevation: < 25 m	Soil series: Tarboro sandy loam and loamy sand		
Slope per cent: < 1.0	Top soil depth: 20 cm		
Slope aspect: 317° NW	Soil depth: > 1.5 m		
Slope shape: linear convex	Drainage class: very well to excessively well drained		
Profile position: shoulder			
Agonomic Management			
Present crop: corn DK 657			
Rotation: corn/sweet potato/cotton			
Nutrient inputs: 10-34-0 @ planting; 30% UAN @ lay by			
Current season soil amendments: none			
Herbicides: metolachlor + atrazine (Bicep) - pre; ametryn (Evik) - post directed			
Insecticides: chlorpyrifos (Lorsban 15G) @ planting			
Tillage: no-till corn/strip till cotton/conventional bedded sweet potato			
Cover crop: rye			

Analysis

Soil from the non-traffic position was submitted to the NCDA labs for diagnostic testing and nematode assay. Soil pH, soluble salts (EC) and mineral nitrogen (NO_3^- -N) were determined on site using an Orion model 290A pH meter, Hanna Instruments DiST 4 dissolved solids tester, and semi-quantitative Aqua-Check colorimetric nitrate test strips, respectively (Figure 4). In addition, soil NO_3^- -N was determined quantitatively in the laboratory by extracting with 1 molar concentration potassium chloride. The extractant was then analyzed using a Lachat Quick Chem 8000 flow injection analyzer.

Soil bulk density was determined with an Uhland drive hammer, tube, and core assembly (figure 5). Derived physical indicators such as porosity and water-filled pore space were obtained by standardized equations using bulk density values.

Visual descriptors of soil characteristics were reported following the Munsell Soil Color Charts system (2000 edition) and the National Soil Survey Center's Field Book for Describing and Sampling Soils version 1.1 (Natural Resources Conservation Service, Lincoln, Nebraska 1998).



Figure 3. Sampling 45 cm deep using a custom drive hammer and steel tube. Obtaining a representative sample is critical for precise diagnostic inference.



Figure 4. Measurement of pH and soluble salts can be determined quickly in the field.



Figure 5. Interns learned to measure soil bulk density and interpret the results. Information derived from bulk density provides insight about the physical condition of the soil with respect to entry and transfer of air, moisture, and heat.

Results

Since pH exerts a profound influence on soil chemistry, this was one of the first tests conducted on site. The acute acidity problem was detected immediately: soil pH ranged from 4.5 to 4.6 in the surface 15 cm through 30 cm deep at the Pr site (Table 2). Low base saturation, 29%, associated with high acidity, was also detected in the surface 30 cm. The target pH for mineral soils in North Carolina is 6.0. As pH drops below 5.0 the quantity of soluble trivalent aluminum (Al^{+3}) increases dramatically in the soil solution. Many

agronomically important crops are sensitive to Al^{+3} even in low concentrations, while elevated Al^{+3} limits root growth as well as nutrient absorption. Manganese (Mn) is a plant-essential micronutrient, but high soluble Mn^{+2} in the root zone can also be toxic to plants. Given that Al^{+3} was probably present and Mn^{+2} concentration in the soil solution was also high, it is questionable how well corn roots were functioning.

Table 2. Soil chemical characterization in the untrafficked interrow.

Depth (cm)	pH (H ₂ O)		CEC (meq/100 ³)		Base saturation (%)		EC (ds/m)		NO ₃ ⁻ - N (ppm)	
	Pr site	Non-Pr site	Pr site	Non-Pr site	Pr site	Non-Pr site	Pr site	Non-Pr site	Pr site	Non-Pr site
0-15	4.5	5.1	2.1	3.1	29	55	0.14	0.08	15.5	6.9
15-30	4.6	5.2	1.7	2.6	29	58	0.11	0.05	4.2	2.8
30-45	5.0	5.3	2.0	2.8	60	71	0.12	0.10	7.5	5.2

Table 3. NCDA soil nutrient analysis.

Nutrient (ppm)	Depth (cm)	Pr site		Non-Pr site		NO ₃ ⁻ - N (ppm)	
			Sufficiency rating		Sufficiency rating	Pr site	Non-Pr site
Phosphorus (P)	0-15	239	V. high	220	V. high	15.5	6.9
	15-30	161	V. high	113	High	4.2	2.8
	30-45	62	High	28	Low	7.5	5.2
Potassium (K)	0-15	86	Medium	152	High		
	15-30	88	Medium	152	High		
	30-45	149	High	152	High		
Calcium (Ca)	0-15	58	Low	205	Medium		
	15-30	44	Low	166	Medium		
	30-45	116	Medium	224	Medium		
Magnesium (Mg)	0-15	10	Low	30	Low		
	15-30	7	Low	37	Medium		
	30-45	26	Low	54	Medium		
Sulfur (S)	0-15	1.4	Medium	1.3	Medium		
	15-30	1.2	Medium	1.2	Medium		
	30-45	0.84	Low	2.2	Medium		
Manganese (Mn)	0-15	28	V. high	50	V. high		
	15-30	48	V. high	42	V. high		
	30-45	41	V. high	33	V. high		
Zinc (Zn)	0-15	1.3	Medium	2.5	Medium		
	15-30	1.1	Medium	1.8	Medium		
	30-45	2.0	Medium	0.68	Low		
Copper (Cu)	0-15	1.1	Medium	1.1	Medium		
	15-30	1.1	Medium	1.0	Medium		
	30-45	1.2	Medium	1.0	Medium		

Cation exchange capacity (CEC: Table 2) was low, but this is normal for sandy southeastern Atlantic coastal plain soils.

Soluble salts and NO_3^- -N were nearly two-fold greater at the Pr site as compared to the non-Pr site (Table 2). These results agree with our understanding that fertilizer N uptake is often depressed in low pH soil due to impaired root growth and function. Accumulation of fertilizer NO_3^- -N in the surface 15 cm provided further evidence of reduced root N uptake by plants growing in acid, high Al^{+3} and Mn^{+2} soil. As noted in Table 2, soil at non-Pr site was also strongly acid

but less acid compared to the Pr site. Consequently, soils at both of the sampled sites in CAR-20 were found low in calcium (Ca) and magnesium (Mg). Sulfur (S) was also marginal at both sites (Table 3). Deficiencies of Ca, Mg, S, coupled with excessive accumulation of Mn in the corn leaf tissue were mainly responsible for the visual symptoms of plant stunting, yellowing, and interveinal chlorosis apparent at the Pr site (Figure 6).

Table 4. Comparison of the critical range for corn leaf tissue nutrient concentrations in the most recently matured (MRM) leaf.

Nutrient	Critical range MRM leaf	MRM tissue analysis Pr site	Comment	Non-Pr site	Comment
— % —					
Nitrogen	3.0 - 4.0	4.07	High	3.48	Sufficient
Phosphorus	0.30 - 0.50	0.30	Sufficient	0.21	Low
Potassium	2.0 - 3.0	2.61	Sufficient	1.57	Low
Calcium	0.25 - 0.80	0.23	Low	0.27	Sufficient
Magnesium	0.15 - 0.60	0.07*	Deficient	0.08*	Deficient
Sulfur	0.15 - 0.40	0.09*	Deficient	0.12	Low
— ppm —					
Boron	5 - 25	8.4	Sufficient	6.8	Sufficient
Copper	5 - 25	5.6	Sufficient	8.1	Sufficient
Iron	30 - 250	83.7	Sufficient	79.5	Sufficient
Manganese	20 - 150	287 (528)**	Excessive	52.1	Sufficient
Zinc	20 - 70	60.3	Sufficient	38.8	Sufficient

* Most limiting

** Number in parentheses is corn leaf tissue Mn concentration two weeks after initial sampling



Figure 6. Corn plant showing symptoms of yellowing and interveinal chlorosis associated with nutrient deficiencies of Mg and S.

Plant stunting and yellowing are classic symptoms of 'acid soil infertility'. On the other hand, symptoms of acute toxicity and/or plant mortality at the Pr site were probably the result of contact with the post-directed herbicide ametryn or 30% N solution, or both, applied at lay-by as plants were severely stunted at this point and, the foliage more susceptible to stray contact with agrochemicals. Nematode assays were negative at both sites.

Manganese concentration in the corn leaf tissue from the Pr-site was very high and increased nearly two-fold after two weeks (Table 4). In strongly acid soil, excessive Mn accumulation occurs through the root system which is toxic to plants at high concentration. At first glance it may appear odd that corn leaf tissue N content was above the sufficiency level at the Pr-site when test results showed an accumulation of fertilizer NO_3^- -N in the surface 15 cm. This apparent anomaly may be explained by the 'concentration effect' brought about by plant stress, particularly moisture, due to impaired root function as compared to healthy, unstressed, rapidly growing corn plants.

A comparison of semi-quantitative (Aqua-check) vs quantitative (flow injection analysis) estimates of NO_3^- -N in the surface 15 cm showed that the semi-quantitative method significantly underestimated soil NO_3^- -N at both sample sites (Figure 7). This points up the fact that soil test kits based on semi-quantitative tests for N, P, K and other nutrients may be useful indicators when accuracy is not critical but are inappropriate for quantitative testing.

Examination of subsurface soil borings revealed a coarse-textured profile to 150 cm deep at the Pr-site. In contrast, the non-Pr site had a distinct horizon of silt and clay enrichment 30-70 cm deep (Figures 8 and 9). Compact, fine textured soil horizons typically impede drainage and may explain the accelerated rate of leaching and consequent lower pH in the Pr-site compared to the non-Pr site. Furthermore, coarse textured, low organic matter soils have lower cation exchange capacity, limiting their ability to supply plant essential nutrients like Ca, Mg, and K.

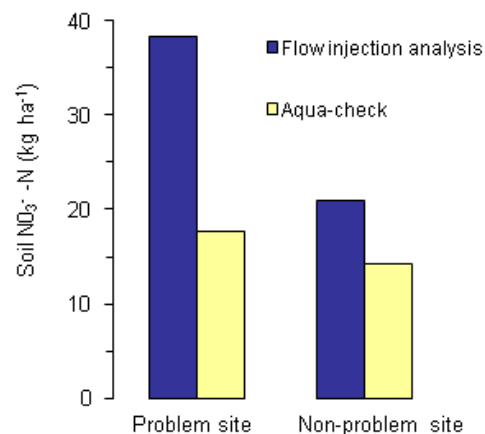


Figure 7. Comparison of quantitative (flow injection analysis) and semi-quantitative (Aqua-check) estimates of soil NO_3^- -N in the surface 15 cm at two sampling sites.

Table 5. Soil physical properties.

Index	Depth (cm)	Pr site		Non-pr site	
		Non-traffic Inter-row	Trafficked Inter-row	Non-traffic Inter-row	Trafficked Inter-row
Bulk density (g/cm^3)	0 - 7.5	1.56	1.65	1.33	1.68
	7.5 - 15	1.67	1.69	1.84	1.66
Porosity (%)	0 - 7.5	41.3	37.6	49.9	36.4
	7.5 - 15	37.0	36.3	30.7	37.3
Water-filled pore space (%)	0 - 7.5	38.9	53.6	44.0	68.3
	7.5 - 15	47.7	54.7	14.7	57.8

Our subsurface investigations testify that soils can be quite variable in terms of their physical and chemical characteristics even though appearing indistinct at the surface. Abrupt changes in subsurface soil texture may occur over short distances even within the same diagnostic mapping unit. In this case, excessive leaching in a Tarboro loamy sand was the culprit leading to locally depressed pH at the Pr-site. We noted several other locations within the CAR-20 unit exhibiting the same crop symptoms observed at the Pr site; we speculate that variability in subsurface texture is probably the causal factor accelerating soil acidity in those areas as well.

Conclusion

Agronomic problem solving often involves consideration of many agents reaching beyond the obvious suspects of human error or natural coincidence. It would have been easy to conclude for instance, that symptoms of acute corn plant toxicity and/or mortality were the result of contact with herbicide and/or soluble fertilizer. However, we learned much more than this by digging deeper and leaving no stone unturned in our investigation. Using a multi-faceted approach gathering information about the site, soil (surface and subsurface), plant (shoots and roots) as well as historic management practices enables the investigator to trouble shoot agronomic problems with greater precision. It may also lead to new insights about the complex soil-plant-atmosphere realm that drives agricultural productivity.

Parent material: coastal plain alluvium
Moisture status: moist throughout profile
USDA taxonomic order: Entisol

Depth (cm)	Description
0 - 20	10YR 5/2 grayish brown very friable medium loamy sand (>80% sand) + few gravel (<1%); weak granular structure
20 - 38	10YR 5/6 yellowish brown very friable medium loamy sand (>80 sand) + few gravel (<1%); very weak medium granular structure
38 - 76	7.5YR 5/8 strong brown medium sand + few gravel (<2%); loose single grain structure
76 - 107	7.5YR 5/8 strong brown medium to coarse sand + gravel (~8%); loose single grain structure
107 - 150	7.5YR 5/8 strong brown to 10YR 6/6 brownish yellow medium to coarse sand + gravel (~20%); loose single grain structure

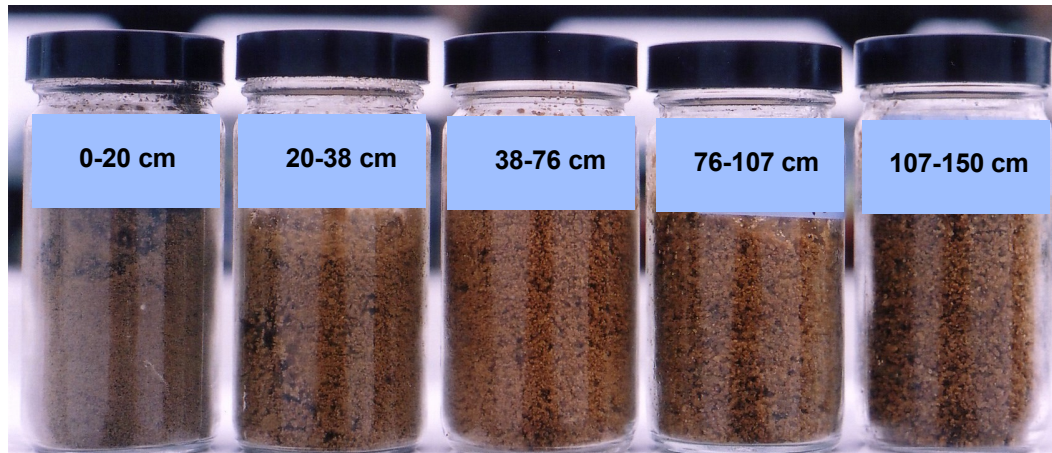


Figure 8. Soil samples recovered from 0-150 cm deep at the problem site. Note the uniform coarse texture throughout the sampled profile in this Tarboro loamy sand.

Parent material: coastal plain alluvium
USDA taxonomic order: Entisol
Moisture status: moist throughout profile

Depth (cm)	Description
0 - 28	10YR 4/3 brown very friable medium to fine loamy sand; weak medium granular structure
28- 70	2.5YR 4/6 red firm silty clay loam; coarse moderate sub-angular blocky structure
70- 102	7.5YR 5/8 strong brown loose medium sand + few gravel (<1%); loose single grain structure
102-150	7.5YR 6/8 reddish yellow coarse sand + few gravel (~5%); loose single grain structure

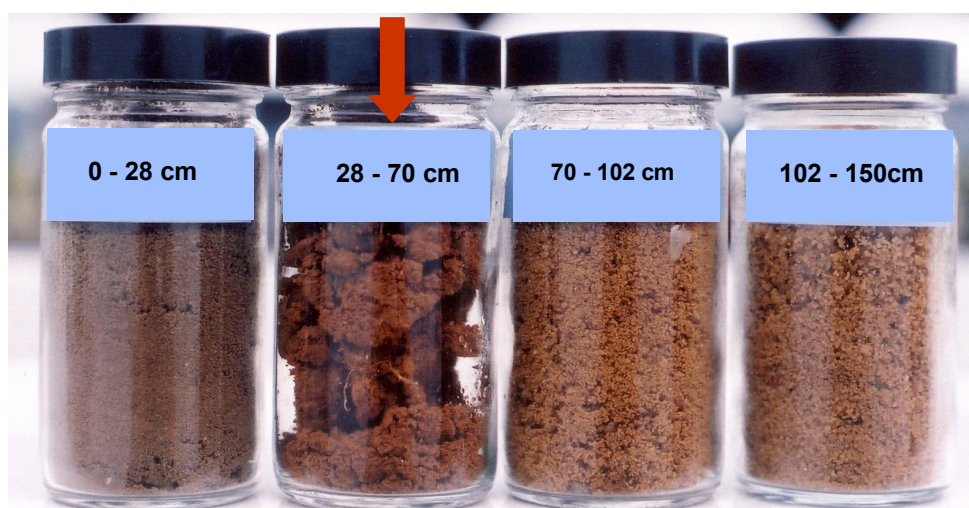


Figure 9. Soil samples recovered from 0-150 cm depth at the non-problem site. Red arrow points to distinct firm silty clay loam horizon differentiating this soil from that found at the problem site. Contrasting soils in Figures 8 and 9 were proximate in the same mapping unit but have different subsurface characteristics.

Following are some recommendations and conclusions submitted from this investigation.

- Overall, the soil in SARE CAR-20 unit is lime deficient. Since there would be little benefit liming soil in the current season, soil from this plot should be submitted in fall to NCDA labs for recommendations aimed at correcting this problem in 2004 and thereon.
- Tarboro loamy sands are very well to excessively well drained soils. Therefore, annual soil testing should be conducted to assess pH and nutrient levels. More frequent liming may be needed for Tarboro loamy sands than is presently practiced.
- Variable subsurface soil textures found in CAR-20 indicate that disjoint areas of poor growth may persist without precision management. This is the fundamental concern of variable-rate technology (VRT) emerging within the field of precision agriculture. However, precision management is not an option at this time as VRT has not been implemented on the farm. Coarse textured, very well to excessively well drained North Carolina coastal plain soils have limited nutrient and water holding capacity. A program integrating intensive cover crop use, returning crop residue, and increasing soil organic matter should conserve moisture while enhancing nutrient holding capacity.

Further Reading

Bennett, W.F., ed. 1993. *Nutrient Deficiencies and Toxicities in Crop Plants*. American Phytopathological Society. St Paul, Mn. Excellent publication offered by the APS. This volume includes many distinct photos of plant symptoms caused by biotic and abiotic factors.

Food and Agriculture Organization of the United Nations. 1984. *Fertilizer and Plant Nutrition Guide*. FAO Fertilizer and Plant Nutrition Bulletin No. 9. Food and Agriculture Organization. Rome, Italy. Over the years the FAO has published many excellent technical bulletins that, unfortunately, have not seen a very wide distribution. This title is dated but remains a solid reference.

Sprague, P.F., ed. 1964. *Hunger Signs in Crops*, 3rd ed. David McKay and Company. New York, NY. Originally published jointly by the American Society of Agronomy and the National Fertilizer Association in 1941, this out-of-print classic remains one of the best visual guides.

